

Endophytic Microbes: As a Source of Antibiotics and Anticancer Agents



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Abstract

Endophytes are the microorganisms which reside inside the plant tissues. They are the great source of biologically active compounds. They can also be the reason for a special property in the plant. Several biologically active molecules have been reported from endophytes. They can be good source of therapeutic molecules, drugs against serious diseases, plant growth promoting molecules, antioxidants and many more. Endophytes are now being considered as treasurer of beneficial compounds. Looking at the limitations regarding isolation of biologically active molecules from plants, endophytes are gaining more attention for the same purpose. A lot of research is being done on endophytic bacteria and fungi in order to procure good amount of beneficial compounds. It is expected that almost all plants possess good number of endophytes. In order to face the challenges in health and agriculture sectors more investigation is required to harness the hidden potential in endophytes.

Keywords: Endophytes, biologically active compounds, bacteria, fungi
Introduction

Natural products obtained from endophytic microbes have great potential in the pharmaceutical and agrochemical industries. A significant number of interesting molecules such as steroids, alkaloids, isocoumarins, quinones, flavonoids, phenylpropanoids, lignans, peptides, phenolics, aliphatic, and volatile organic compounds, many of them biologically active have been obtained from endophytic microbes. (Tan and Zou, 2001; Gunatilaka, 2006; Zhang *et al.*, 2006). According to Strobel (2002) endophytic fungi are a promising source of novel compounds. It has been estimated that about 51% of biologically active substances from fungal endophytes were previously unknown. Endophytes also produce extracellular hydrolyases which provides resistance mechanism against plant invasion and some of the extracellular enzymes like cellulases, proteinase, lipases and esterases. The actions of these enzymes support the hypothesis of co-evolution between endophytes and their respective hosts. Many secondary metabolites produced by fungal endophytes are larger than that of any other endophytic microbes (Zhang *et al.*, 2006). Endophyte opens up with new and different areas for the biotechnological exploitations. Crude extracts from culture broth of endophytes shows antibacterial, antifungal, antiviral, anti-inflammatory and antitumor activities (Silva *et al.*, 2007). It has been reported that plant derived natural product is not up to the desired level, as it is produced at a specific development stage or under specific environmental condition, stress or nutrient availability. Moreover, plants may take many years to reach a suitable growth phase for product accumulation. Hence, considering these limitations, endophytes are the alternative source of bioactive natural compounds. There are number of endophytes which are the members of common soil bacteria genera like *Pseudomonas*, *Burkholderia* and *Bacillus* species (Lodewyckx *et al.*, 2002). Wide range of biological active compounds have been isolated from endophytic microorganisms. These microorganisms are still remaining a relatively untapped source of natural products. Current interest in natural bioactive compounds from endophytes especially endophytic fungi is evident from the no of review articles which have been appeared in recent literatures (Hasegawa *et al.*, 2006; Zhang *et al.*, 2006; Gunatilaka 2006; Guo *et al.*, 2008; Staniek *et al.*, 2008; Ryan *et al.*, 2008).

Endophytic Microorganisms as a Source of Antibiotics

Natural products from endophytic microbes have been observed to inhibit or kill a wide variety of harmful disease causing agents including phytopathogens, bacteria, fungi, viruses and protozoans that affect humans

and animals. *Cryptosporiopsi quercina* was isolated as an endophyte from *Tripterygium wilfordii*, a medicinal plant native to Eurasia (Strobel et al., 1999). *C. quercina* showed an excellent antifungal activity against some important fungal pathogens such as *Candida albicans* and *Trichophyton* sp. Reports suggested that a unique peptide antimycotic, termedas cryptocandin, was isolated and characterized from *C. quercina* (Strobel et al., 1999). The antibiotic ecomycins are produced by *Pseudomonas viridiflava*, member of a group of plant-associated fluorescent bacteria (Miller et al., 1998). The ecomycins are very active against human-pathogenic fungi as *Cryptococcus neoformans* and *Candida albicans*. Colletotric acid, a metabolite of *Colletotrichum gloeosporioides*, an endophytic fungus in *Artemisia mongolica*, displays antimicrobial activity against bacteria and fungus *Helminthosporium sativum* (Zou et al., 2000).

In addition to plants such as *A. annua* producing antimalarial compounds, some endophytes have also shown strong activity against protozoan diseases as well. Wide-spectrum antibiotics are produced by *Streptomyces* sp. strain NRRL 30562, an endophyte in *Kennedia nigricans*. These antibiotics, called munumbicins, possess widely differing biological activities, depending on the target organism. The munumbicins demonstrate activity against *Bacillus anthracis* which is gram positive bacteriaum and multidrug-resistant *Mycobacterium tuberculosis*, *Plasmodium falciparum* and other drug-resistant bacteria (Castillo et al., 2002). The endophytic *Chloridium* sp. from *A. indica* produces Javanicin which is known to be highly effective and active against *Pseudomonas* sp. (Kharwar et al., 2008). A strain of *P. microspora* was isolated from the endangered tree *Torreya taxifolia* which produces several bioactive compounds having antifungal activity, including pestaloside, an aromatic β glucoside, and two pyrones such as: pestalopyrone and hydroxypestalopyrone. These products possess phytotoxic properties (Lee et al., 1995). According to Strobel et al., (1996 & 2002) *Pestalotiopsis microspora* is a common rainforest endophyte which has an enormous biochemical diversity which exists in this endophytic fungus and produces many secondary metabolites. One such secondary metabolite is ambuic acid, which is an antifungal agent and has been recently described from several isolates of *P. microspora* (Li et al., 2001). According to Horn et al., 1995, phomopsichalasin, a metabolite obtained from an endophyte of *Phomopsis* sp., represents the first cytochalasin type compound with a three ring system replacing the cytochalasin macrolide ring. Phomopsichalasin metabolite mainly exhibits antibacterial activity against *Salmonella enterica* serovar Gallinarum, *Bacillus subtilis* and *Staphylococcus aureus*. It shows moderate activity against the yeast *Candida tropicalis*. Colletotric acid, a metabolite of *Colletotrichum gloeosporioides*, an endophytic fungus in *Artemisia mogolica*, displays antimicrobial activity against bacteria as well as against the fungus *Helminthosporium sativum* (Zou et

al., 2000). Another *Colletotrichum* sp., isolated from *Artemisia annua*, produces bioactive metabolites that showed varied antimicrobial activity. The *Colletotrichum* sp. found in *A. annua* produced not only metabolites with activity against human pathogenic fungi and bacteria but they were also metabolites that were fungistatic to plant pathogenic fungi (Liu et al., 2000).

Endophytic Microorganisms as Source of Anticancerous Agent

According to Suffness, 1995, Paclitaxel is a highly functionalized diterpenoid which is found in each of the world's yew (*Taxus*) species. Paclitaxel and its derivatives represent the first major groups of anticancerous compounds that are produced by endophytes. It has been reported that a novel paclitaxel-producing endophytic fungus, *Taxomyces andreanae*, was discovered in *Taxus brevifolia* (Strobel et al., 1993). Most commonly found endophytes of the world's yews are *Pestalotiopsis* sp. (Strobel, 2002). Endophytes of *Taxus wallichiana* yielded *P. microspora*, and a preliminary monoclonal antibody test indicated that it might produce paclitaxel (Strobel et al., 1996). This clearly showed that endophytes residing in plants other than *Taxus* sp. were producing paclitaxel. Chaetopyranin is a benzaldehyde derivative isolated from the endophytic fungus *Chaetomium globosum* associated with the marine red alga *Polysiphonia urceolata*. Chaetopyranin exhibited moderate to weak cytotoxic activities against three human tumor cell lines HMEC (human microvascular endothelial cells), SMMC-7721 (hepatocellular carcinoma cells) and A54 (human lung epithelial cells) (Wang et al., 2004). Some of the most potent plant-derived antileukemic alkaloids have also been reported from endophytic fungi. Another important compound is camptothecin, which is a chemotherapeutic agent obtained from medicinal plants and was firstly isolated from *Camptotheca acuminata* (Wall et al., 1966). It has an unusual efficacy against lung, ovarian, and uterine cancers. Camptothecin basically acts as strong inhibitors of topoisomerase I by trapping the cleavage DNA-topoisomerase I complex (Torck and Pinkas, 1996). The endophytes *Rhinocladiella* sp. from *Tripterygium wilfordii* produces three new chalasins that inhibit cell division of colon and ovarian tumor cell lines (Wagenaar et al., 2000). Asparaginase an antileukaemic agent is produced from a variety of microbial sources including fungi (Sarquis et al., 2004), yeasts (Ferrara et al., 2006) and bacteria (Geckil and Gencer, 2004). Theantana et al., 2009 produced asparaginase from wild medicinal plants in Thailand namely, *Adenanthera microisperma*, *Betula alnoides*, *Cassia alata*, *Houttuynia cordata* and *Hiptage benghalensis*. Asparagine amino acid is a nutritional requirement of both normal cells and cancer cells. Normal cells have the ability to produce enzyme asparagine synthetase, which can synthesize asparagine from aspartic acid, whereas in cancer cells this enzyme is present in a very low level (Nakamura et al., 1999). Very low levels of these nonessential amino acid asparagines will only affect

the viability of abnormal cells as these cells have abnormal requirements for asparagine (Haley *et al.*, 1961; Mitchell *et al.*, 1994).

Table 1: Natural Products Reported From Endophytes of Different Host Plants (Ref. Shodhganga)

Host plant	Endophytic fungi	Bioactive compound	Activity	References
<i>Garcinia dulcis</i>	<i>Phomopsis</i> sp.	Phomoenamide	Antimicrobial	Rukachaisirikul <i>et al.</i> , 2008. <i>Phytochemistry</i> 69: 783-787
<i>Artemisia mangolica</i>	<i>Colletotrichum gloesporides</i>	Colletotric acid	Antifungal & Antibacterial	Zou <i>et al.</i> , 2000. <i>J Nat. Prod</i> 63:602-604
<i>Cinnamomum zeylanicum</i>	<i>Muscodor albus</i>	Volatile organic compounds	Antifungal/ Antibacterial	Ezra <i>et al.</i> , 2004 <i>Microbiology</i> . 150:4023-4031
<i>Taxus brevifolia</i>	<i>Taxomyces andreannae</i>	Paclitaxel	Anticancer	Stierle <i>et al.</i> , 1993. <i>Science</i> 260:214-216
<i>Taxus wallichiana</i>	<i>Pestalopsis microspora</i>	Paclitaxel	Anticancer	Strobel, 1996. <i>Journal of Industrial Microbiology</i> 142:435-440
<i>Taxus cuspidate</i>	<i>Periconia</i> sp	Periconicins A&B	Antibacterial	Kim <i>et al.</i> , 2004. <i>J.Nat Prod</i> 67:448-450
<i>Taxus chinensis</i>	<i>Fusarium solani</i>	Tax3	Anticancer	Strobel 1996. <i>Microbiology</i> 142:435-440
<i>Tripterygium wilfordii</i>	<i>Rhinodadiella species</i>	Cytochalasin	Antibiotic	Wagenaar <i>et al.</i> , 2000 <i>J.Nat Prod</i> . 63:1692-1695
<i>Terminalia morobensis</i>	<i>Pestalotiopsis microspora</i>	Pestacin & Isopestcin	Antioxidant & antimicrobial	Strobel <i>et al.</i> , 2002 <i>Crit Rev Biotechnol</i> 22:315-333
Woody plant	<i>Nodulisporium</i> sp	Nodulisporic acid	Insecticidal	Findlay <i>et al.</i> , 1997. <i>J.Amchem Soc.</i> 119:8809-8816
<i>Paullinia Paullinioides</i>	<i>Muscodor 4TJKKKKTTTvitigen</i>	Naphthalene	Insecticidal	Daisy <i>et al.</i> , 2002. <i>Mycotaxon</i> . 84:39-50
<i>Tripterygium wilfordii</i>	<i>Fusarium subglutians</i>	Subglutinol A&B	Immunosuppressive	Lee <i>et al.</i> , 1995. <i>J. Org.Chem</i> , 60:7076-7077
<i>Eucryphia cordifolia</i>	<i>Muscodor albus</i> & <i>Gliocladium</i> sp	Volatile organic compound	Antibiotic	Stinson <i>et al.</i> , 2003 <i>Plant science</i> 165:913-922
Xylopia aromatic	<i>Periconia atropurpurea</i>	Perconian B	Cytotoxic activity	Teles <i>et al.</i> , 2006, 67:2686-2690
<i>Taxodium distichum</i>	<i>Pestalotiopsis microspora</i>	Taxol	Anticancer	Li <i>et al.</i> , 1996. <i>Microbiology</i> 142:2223-2226
<i>Torreya grandiflora</i>	<i>Perconia</i> species	Taxol	Anticancer	Li <i>et al.</i> , 1998. <i>J.Ind Microbiol Biotechnol</i> 20:259-264
<i>Wallemia nobilis</i>	<i>Pestalotiopsis guipini</i>	Taxol	Anticancer	Strobel <i>et al.</i> , 1997. <i>Aust J. Bot</i> 45:1073-1082

References

1. Tan RX, Zou WX., *Endophytes: a rich source of functional metabolites: Nat Prod Rep.*, August 2001,18(4):448-59.
2. A Leslie Gunatilaka, *Natural Products from Plant-associated Microorganisms: Distribution, Structural Diversity, Bioactivity, and Implications of Their Occurrence*, *J Nat Prod.* 2006 Mar; 69(3): 509–526.
3. JT Zhang, W Ru, B Li, *Relationships between vegetation and climate on the Loess Plateau in China*, *Folia Geobotanica*, 2006, 41, pp. 151-163
4. Strobel GA., *Rainforest endophytes and bioactive products*, *Crit Rev Biotechnol.* 2002; 22(4):315-33
5. E Dilip de Silva, Pamoda Bhasini Ratnaweera, *Endophytic Fungi: A Remarkable Source of Biologically Active Secondary Metabolites*, In book: *Endophytes: Crop Productivity and Protection*, November 2017.
6. Cindy Lodewyckx, Jaco Vangronsveld, Fiona Porteous, Edward R. B. Moore, Safieh Taghavi, Max Mezgeay, Daniel van der Lelie *Endophytic Bacteria and Their Potential Applications*, *Critical Reviews in Plant Sciences*, November 2002, 21(6):583-606.
7. Sachiko Hasegawa, Akane Meguro, Masafumi Shimizu, Tomio Nishimura, Hitoshi Kunoh, *Endophytic Actinomycetes and Their Interactions with Host Plants*, *Actinomycetologica*, January 2006, 20(2):72-81.
8. Zhang HW, Song YC, Tan RX, *Biology and chemistry of endophytes*, *Nat Prod Rep.*, 2006 Oct, 23(5):753-71.
9. BY Guo, Y Wang, Xiulan Sun, K Tang, *Bioactive natural products from endophytes: A review*, *Prikladnaia biokhimiia i mikrobiologii*, March 2008, 44(2):153-8
10. Agata Staniek, Herman J. Woerdenbag, Oliver Kayser, *Endophytes: Exploiting biodiversity for*

- the improvement of natural product-based drug discovery, Journal of Plant Interactions, June 2008, 3(2):75-93*
11. Ryan RP, Germaine K, Franks A, Ryan DJ, Dowling DN., *Bacterial endophytes: recent developments and applications. FEMS Microbiol Lett.* 2008 Jan; 278(1):1-9
 12. Strobel GA, Li JY, Sugawara F, Koshino H, Harper J, Hess WM. *Oocydin A.. A chlorinated macrocyclic lactone with potent anti-oomycete activity from Serratia marcescens* *Microbiol*, 1999, 145: 3557-64.
 13. Miller, CM, Miller, RV, Garton-Kenny, D, Redgrave, B, Sears, J, Condon, MM, et al. *Ecomycins, unique antimycotics from Pseudomonas viridisflava*. *J Appl Microbiol*, 1998, 84: 937- 44.
 14. Zou, WX, Meng, JC, Lu, H, Chen, G X, Shi, GX, Zhang, T Y, & Tan, RXM *Metabolites of Colletotrichum gloeosporioides, an endophytic fungus in Artemisia mongolica*. *Journal of Natural Products*, 2000, 63, 1529-1530.
 15. Castillo U, Strobel GA, Ford EJ, et al., *Munumbicins, wide spectrum antibiotics produced by Streptomyces munumbi, endophytic on Kennedia nigriscans*. *Microbiology*, 2002, 148: 2675-2685.
 16. Ravindra N Kharwar, VC Verma, G Strobel, David Ezra, *The endophytic fungal complex of Catharanthus roseus (L.) G. Don*, *Current Science*, July 2008, 95(2):228-233
 17. Lee, JC, Lobokovsky, NB, Plam, NB, Strobel, GA and Clardy, JC. *Subglutinol A and B: immunosuppressive compounds from the endophytic fungus Fusarium subglutinans*. *Journal of Organic Chemistry*, 1995, 60, 7076-7077.
 18. Strobel, GA. *Microbial gifts from the rainforest*. *Canadian Journal of Phytopathology*, 2002, 24, 14-20.
 19. Liu, CH, Zou, WZ, Lu, H and Tan, RX *Antifungal activity of Artemisia annua endophyte cultures against phytopathogenic fungi*. *Journal of Biotechnology*, 2001, 88:277-282.
 20. Horn WS, Monique SJ, Simmonds RE, Schwartz WM, Blaney P *Phomopsichalasin, a Novel Antimicrobial Agent from an Endophytic Phomopsis sp*, *Tetrahedron*, April 1995, 51(14):3969-3978.
 21. Gary Strobel, Xianshu Yang, It Joe Sears, Robert Kramer, Rajinder S. Sidhu, WM Hess, *Taxol from Pestalotiopsis microspora, an endophytic fungus of Taxus wallachiana*, *Microbiology* 1996, 142, 43 5-440.
 22. Strobel, G A *Endophytes as sources of bioactive products*. *Microbes and Infection*, June 2003 5(6):535-44.
 23. Ji-Guang Wei, Tong Xu1, Liang-Dong Guo, Ai-Rong Liu1, Ying Zhang and Xiu-Hu Pan, *Endophytic Pestalotiopsis species associated with plants of Podocarpaceae, Theaceae and Taxaceae in southern China*, *Fungal Diversity*, 2007, 24: 55-74.
 24. Wall, M E, MC Wani, CE Cooke, K.T. Palmer, A.T. McPhail and G.A. Sim. *J. Am. Chem. Soc.*, 1996, 88: 3888-3890
 25. M Torck, M Pinkas, *Camptotheycin and derivatives: A new class of antitumor agents*, *Journal de pharmacie de Belgique*, July 1996, 51(4):200-7
 26. Wagenaar, M. M., Corwin J, Strobel G., & Clardy J, *Three new cytochalasins produced by an endophytic fungus in the genus Rhinocladiella*. *Journal of Natural Products*, 2000, 63, 1692-1695.
 27. MID Sarquis , EMM Oliveira, AS Santos, GL da Costa *Production of L-asparaginase by filamentous fungi*, *Memórias do Instituto Oswaldo Cruz*, August 2004, 99(5):489-492
 28. Ferrara MA, Severino NMB, et al., *Asparaginase production by a recombinant Pichia pastoris strain harbouring Saccharomyces cerevisiae ASP3 gene*. *Enzyme Microb. Tech.*, 2006, 39 : 1457-1463
 29. Hikmet Geckil, Salih Gencer, *Production of L-asparaginase in Enterobacter aerogenes expressing Vitreoscilla hemoglobin for efficient oxygen uptake*, March 2004, *Applied Microbiology and Biotechnology*, March 2004, 63(6):691-7
 30. Nakamura CT, Wilkinson R et al., *Pancreatitis and peritonitis following therapy with L-asparaginase*, *Int. Pediatr. Res.*, 1999, 14: 25-27.
 31. Teerayut Theantana,, Kevin D Hyde, Saisamorn Lumyong, *Asparaginase production by endophytic fungi from Thai medicinal plants: Cytotoxicity properties*. *International Journal of Integrative Biology*, 2009, Vol. 7, No. 1, 1-8.
 32. E E Haley, G A Fischer and A D Welch, *The Requirement for L-Asparagine of Mouse Leukemia Cells L5178Y in Culture*, *Cancer Res* 1961; 21:532-536.
 33. Mitchell I, Hoogendoorn H, Giles AR. *Increased endogenous thrombin generation in children with acute lymphoblastic leukemia: risk of thrombotic complications in L- asparaginase induced antithrombin III deficiency*. *Blood*, 1994, 83: 386-391.
 34. Rukachaisirikul V, Sommart U, Phongpaichit S, Sakayaroj J, Kirtikara K. *Metabolites from the endophytic fungus Phomopsis sp . PSU -D15*. *Phytochemistry*. 2008; 69: 783-787
 35. Ezra D, Hess WM, Strobel GA. *New endophytic isolates of Muscodor albus, a volatile-antibiotic-producing fungus*. *Microbiology*. December 2004, 150, 12:4023-31.
 36. Stierle A, Strobel G and Stierle D, *Taxol and taxane production by Taxomyces andreanae, an endophytic fungus of Pacific yew*; *Science*, 1993, 260 214-216.
 37. GA Strobel, WM Hess, E Ford, RS Sidhu, X Yang, *Taxol from fungal endophytes and the issue of biodiversity*, *Journal of Industrial Microbiology*, November 1996, Volume 17, Issue 5-6, pp 417-423

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38. Kim S, Shin DS, Lee T, Oh KB. *Periconicins, two new fusicoccane diterpenes produced by an endophytic fungus Periconia sp. with antibacterial activity.* J Nat Prod. 2004; 67:448–450.
39. Strobel GA, Rainforest endophytes and bioactive products. Crit. Rev. Biotechnol. 2002; 22(4):315-33.
40. Findlay JA, Li G, Miller JD, Womiloju TO. Can. J. Chem. 2003; 81: 284–292.